

Genotyping of *Echinococcus Granulosus* (Hydatid Cyst) Isolated from Domestic Animals in Kurdistan - Iraq

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Abstract—Hydatid disease is a silent helminthic disease caused by metacestode of dog tapeworm *Echinococcus granulosus*, with medical and economic impact, the recently DNA markers allowed to identify 10 distinct genotypes of *E. granulosus* (G1-G10). The present study is to genotyping of *E. granulosus* (hydatid cyst) isolated from intermediate host. The partial DNA sequences of *mtCOI* gene followed to detect genetic variation of *E. granulosus*. The result of nucleotide sequence alignment with the recorded nucleotide in the NCBI indicated that all samples belong to the sheep strain, and there is a slight variation among sheep strain. A new variant strain was reported and deposited in the GenBank under variant G1 (Kurdistan strain) with accession number G1 (JX878691), sheep strain G1 (JX878689) and (JX878693). The present study concluded that the sheep strain is predominant genotype of *E. granulosus* in Kurdistan - Iraq, also it can be concluded that sheep strain generally responsible for hydatid disease among animal and human.

Index Terms—CO1, *Echinococcus*, Hydatid cyst, Sheep strain.

I. INTRODUCTION

Hydatid cyst is a potential zoonotic disease of man and animals, caused by the metacestode of the dog tapeworm *Echinococcus granulosus* [1]. The cyst can be grown in all organs, whereas the common sites of infection are the liver and lungs [2]. The hydatid cyst has two main forms which differ in pathology and morphology; cystic hydatid disease belongs to *E. granulosus* and alveolar hydatid disease (AHD) which is caused by *Echinococcus multilocularis* [3]. Based on DNA sequence, analysis of the *mtCOI*, dehydrogenase subunit 1 (*NDI*) gene, and *ITS1* 10 genotypes (G1-G10) have been

described in a world isolated from sheep, pigs, cattle, horses, camels, goats, and human [4,5]. The molecular characterization of *E. granulosus* had been studied by many researchers in the different parts of the world [6-14]. The sheep strain is common in human and animal at least seven of these strains have been isolated from humans [15]. The hydatid disease is regarded as a neglected disease, and it remains public health concern because surgical removal of the cyst is the only choice and the recurrence of hydatid cyst still high depending on the operation accuracy and techniques [16,17]. The genotyping of *E. granulosus* has a significant point for control measures and the means of diagnosis [18]. Many studies indicated that Kurdistan Region Government is an endemic area of *E. granulosus* [19-25]. This study carried out to determine the genetic variation of *E. granulosus* and identification of abundance strain of *E. granulosus* in Kurdistan.

II. MATERIALS AND METHODS

A. Sampling

The hydatid cyst was isolated from an animal (10 from sheep, 10 from cattle, and 10 from goats) in Sulaimani modern abattoir. The samples stored separately in a sterile capped containers containing about one volume (v/v) of 70% ethanol at 4°C [26].

B. DNA Extraction

The germinal layer washed 3 times with phosphate buffer saline to remove the ethanol [5]. Modified Tissue extraction kit (Geneaid) followed for DNA extraction, the 0.5 g of each sample was diced to a small piece, then grinded with liquid nitrogen instead of mechanical grinding using micropistol, which provided with the kit, then followed the instruction of the company [27]. The extracted DNA dissolved in 50 µl TE buffer and stored at -20°C until use.

C. Polymerase Chain Reaction (PCR) Process

Nejad and his colleagues [28] protocol were followed to amplify a fragment of *mtCOI* gene, in the 50 µl volume

using a master mix (Cinagene, Iran) contains 10–50 ng of DNA and 50 pmol of each primer forward (5' TTT TTT GGG CAT CCT GAG GTT TAT 3') and reverse (5' TAA AGA AAG AAC ATA ATG AAA ATG 3). The PCR condition sets as follows: An initial stepping at 95°C for 4 min followed by 35 cycles at 94°C for 45 s, 55°C for the 30s, 72°C for 45 s, and 72°C for 7 min, as a final extension step. The result of PCR was detected on 1.5% ethidium bromide-stained agarose gel under UV light and digitally photographed.

D. Sequence Analysis

The GFX PCR DNA product and Gel Band Purification Kit used for PCR product purification (Amersham, UK). The purified PCR product of 12 samples which have a good DNA quantity and purity selected for sequencing in the Center for Research and Biotechnology for Agriculture of Malaya University in Malaysia. Multiple sequence alignments were done using the Clustal W method (Bioedit DNA analysis program).

III. RESULTS

In the present study, the partial *mtCOI* gene of all samples was successfully amplified, and the expected band 443 bps product was detected on 1.5% agarose gel. Amplification of mitochondrial cytochrome subunit 1 (*COI*) gene was detected in all samples, the 369 bp obtained from DNA sequencing for each sample. The multiple sequence alignments carried out with previously published in GenBank of *E. granulosus* under the following accession numbers, Japan sheep strain G1 (U50464), Iran and Jordan sheep strain (JQ250817), G2 (M84662), G3 (M84663), G4 (M84664), G5 (M84665), G6 (M84666), G8 (DQ144021), G10 (AF525457), G1c (AY686565), G1c Argentina (AF458873), and G1 (JN792925), Iran [27,29].

The multiple sequence alignments (Fig. 1) showed that the six isolated samples (sh1, sh4, sh5, sh6, sh7, and go4) were 100% identical to the Japan sheep strain (U504664), whereas five isolates (go1, go3, go5, sh2, and ca2) were 99% identical with sheep strain G1c (AY686565), one sample from goat was differ from the all recorded strains of *E. granulosus* (Table I). The results of the present study deposited in the GenBank as the first record of variant G1 (Kurdistan strain) in Iraq and Kurdistan under the following accession numbers G1 (JX878689), G1 (JX878693), and G1 variant (JX878691).

IV. DISCUSSION

Different molecular markers were described for strain identification and molecular characterization of *E. granulosus* with different efficiency to determine the microvariants and strains [26]. The specific primers for *E. granulosus* strains are reliable for molecular epidemiological study and applicable for the large sample [30]. Many molecular studies support the current study [31] in Iran, [8] in China, [9] in Italy, [10] in India, and [16] in Iraq. They stated that the *COI* has the ability and good efficacy to determine the strains and genetic variations.

The current study revealed that the sheep strain (G1) is the common strain in Kurdistan - Iraq, which has an ability to infect most intermediate hosts including sheep, goats, cattle, and human [28]. The strain identification of *E. granulosus* has a medical important value for control and prevention, whereas the majority of human infection of *E. granulosus* belonged to sheep strain [32]. The high prevalence of sheep strain (G1) and variant G1 (Kurdistan strain) in Sulaimani province and Kurdistan may be due to the intimate commercial contact with Iran; the most imported animals were from Iran which is a concern as an epidemic region of *E. granulosus* [33]. The occurrence of G1 is of significant value for epidemiological and public health importance due to their ability and infectivity to humans.

The DNA sequence of *pmtCOI* gene of sheep strain (G1), Tasmanian sheep strain (G2), and Buffalo strain (G3) is very similar [26], these genotypes G1, G2, and G3 are included in a single cluster, named *E. granulosus* sensu stricto, as suggested by Nakao, *et al.* [34] and previously by other authors [35,36].

V. CONCLUSION

This study concludes that the sheep strain and variant G1 (Kurdistan strain) of *E. granulosus* are mostly responsible for human and animal hydatid cyst in Kurdistan - Iraq, and the Kurdistan - Iraq is the endemic area of *E. granulosus*.

ACKNOWLEDGMENT

The authors would like to thank the CBAR and Directorate General of health/Sulaimani for their cooperation.

TABLE I
THE BASE PAIR CHANGE AND MUTATION OF PMT CO1 SEQUENCE COMPARED WITH THE COMMON SHEEP STRAIN G1 (U50464)

Sample	Intermediate host	Type of cysts	Nucleotide change	Nucleotide position	Genotype	Genebank accession No.
sh1, sh4, sh5, sh6 and sh7	Sheep	Sterile	100% Identity	-	G1	JX878689
go4	Goats	Sterile	100% Identity	-	G1	
go1, go3 and go5	Goats	Sterile	C→T	32	G1c	JX878693
sh2	Sheep	Fertile	C→T	32	G1c	
Ca1	Cattle	Fertile	C→T	32	G1c	
go2	Goats	Fertile	C→T	32	Variant G1 (Kurdistan strain)	JX878691
			T→C	339		

Sh: Sheep, go: Goat, Ca: Cattle

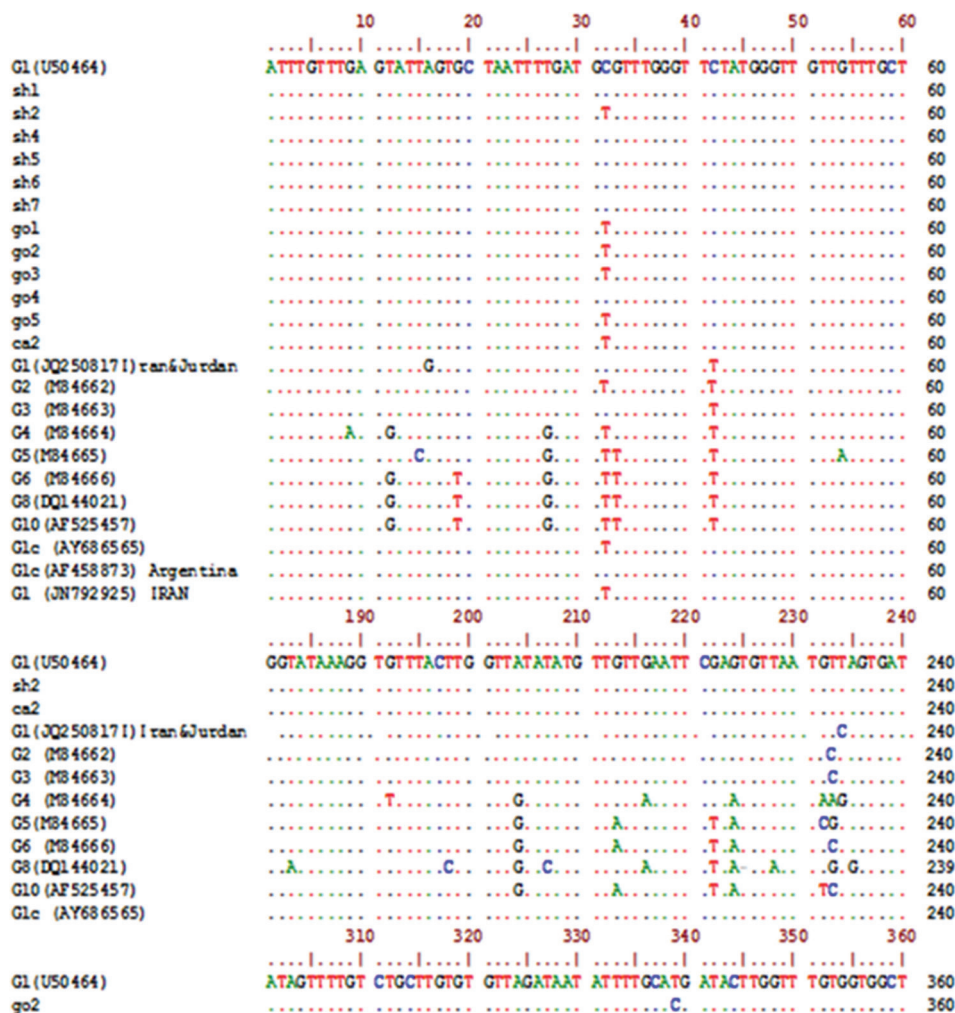


Fig. 1. Representative variable regions of partial nucleotide sequences of 369 bps fragment of the mitochondrial *COI* of hydatid cysts aligned with previously published results.

REFERENCES

- [1] A. R. Satoskar, G. L. Simon, P. J. Hotez and M. Tsuji, *Medical Parasitology*, Texas: Landes Bioscience, 2009.
- [2] E. K. Markell, D. T. John and W. A. Krotoski, *Medical Parasitology*, 8th ed. Philadelphia, PA: WB. Saunder Company, pp. 253-261. 1999.
- [3] World Health Organization Office International des Epizooties. "WHO/OIE manual on echinococcosis in humans and animals," *A Public Health Problem of Global Concern*. Paris, France: World Organization for Animal Health, 2002.
- [4] N. E. Salih and A. L. Jamain, "DNA analysis of *Echinococcus* of human and sheep origin in Ninevah province, Iraq by PCR-RAPD technique," *Journal of Rivista di Parassitology*, vol. 17, no. 3, pp. 221-232, 2001.
- [5] E. Sánchez, O. Cáceres, C. Náquira, D. Garcia and G. Patiño, "Molecular characterization of *Echinococcus granulosus* from Peru by sequencing of the mitochondrial cytochrome C oxidase subunit 1 gene," *Oswaldo Cruz, Rio de Janeiro*, vol. 105, no. 6, 2010.
- [6] L. H. Zhang, J. J. Chal, W. Jiao, Y. Osman and D. P. McManus, "Mitochondrial genomic markers confirm the presence of the camel strain (G6 genotype) of *Echinococcus granulosus* in North-Western China," *Journal of Parasitology*, vol. 116, pp. 29-33, 1998.
- [7] N. Ahmadi and A. Dalimi, "Characterization of *Echinococcus granulosus* isolates from human, sheep, and Camel in Iran," *Journal of Elsevier B.V.*, vol. 6, pp. 85-90. 2006.
- [8] J. M. Bart, M. Abdukader, Y. L. Zhang, R. Y. Lin, Y. H. Wang, M. Nakao and A. Ito. "Genotyping of human cystic echinococcosis in Xinjiang, PR China," *Journal of Parasitology*, vol. 133, pp. 571-579, 2006.
- [9] A. Varcasia, S. Canu, A. Kogkos, A. P. Pipia, A. Scala, G. Garippa and A. Seimenis, "Molecular characterization of *Echinococcus granulosus* in sheep and goats of Peloponnese," *Greece Journal Parasitology Research*, vol. 101, pp. 1135-1139, 2007.
- [10] D. Bhattacharya, A. K. Bera, B. C. Bera, A. Maity and S. K. Das, "Genotypic characterization of Indian cattle, Buffalo and sheep isolate of *E. granulosus*," *Veterinary Parasitology*, vol. 143, pp. 371-374, 2007.
- [11] L. Rinaldi, M. P. Maurelli, F. Capuano, A. G. Perugini, V. Veneziano and S. Cringoli, "Molecular update and cystic echinococcosis in cattle and water buffaloes of southern Italy," *Journal Blackwell Verlag Zoonosis, Public Health*, vol. 55, pp. 119-123, 2008.
- [12] A. Karimi and R. Dianatpour, "Genotypic and phenotypic characterization of *Echinococcus granulosus*," *Boilogy*, vol. 7, no. 4, pp. 757-762, 2008.
- [13] S. Mrad, M. O. Mard, D. Filisetti, M. Mekki, A. Noure, T. Sayadi, E. Candolfi, R. Azaiez, H. Mezhoud and H. Babba, "Molecular identification of *Echinococcus granulosus* in Tunisia: First record of Buffalo strain (G3) in Human and Bovine in the country," *Journal of Open, Veterinary Science*, vol. 4, pp. 27-30, 2010.
- [14] S. Ergin, S. Saribas, P. Yuksel, K. Zengin, K. Midillin, G. Adas, M. Aslan, H. Uysal, R. Caliskan, A. Oner, O. Kucukbasmaci, A. Kaygusuz, M. Torun

- and B. Kocazeybek, "Genotypic characterization of *Echinococcus granulosus* isolated from a human in Turkey," *African Journal of Microbiology Research*, vol. 4, no. 7, pp. 551-555, 2010.
- [15] R. C. Thompson, D. P. McManus, "An etiology parasites and life cycles," In J. Eckert, M. A. Gemmell and F. X. Meslin, Eds. *WHO/OIE Manual on Echinococcosis in Humans and Animals: A Public Health Problem of Global Concern*, Paris: World Organisation for Animal Health, 2002.
- [16] A. A. Hama, W. M. S. Mero and J. M. S. Jubrael, "Molecular identification of *Echinococcus granulosus* (G1) strain in human and animals. *ZUJ Journal*, vol. 1, no.1, pp. 1-6, 2012.
- [17] A. A. Hama, W. M. S. Mero and J. M. S. Jubrael, "Molecular Characterization of *E. granulosus*, First Report of Sheep Strain in Kurdistan-Iraq", 2nd International Conference on Ecological, Environmental and Biological Sciences Bali (Indonesia). EEBS, pp. 41-44, October, 2012.
- [18] M. M. Bajalan, "Prevalence of Echinococcosis in Stray Dogs and Slaughtered Livestock in Kalar District/Sulaimaniyah Province/Kurdistan Iraq," M.Sc. Thesis, College of Veterinary Medicine, University of Baghdad, 2006.
- [19] W. M. S. Mero, J. M. S. Jubrae and A. A. Hama, "Prevalence of hydatid disease among slaughtered animals in Slemani province/Kurdistan-Iraq," *Journal of the University of Zakho*, vol. 2, no. 1, pp. 33-38, 2013.
- [20] H. A. Amin, "Prevalence of Hydatid Cyst in Human and Animal in Sulaimanya and Saed Sadq," M.Sc. Thesis, College of Medicine, University of Sulaimanya, 2007.
- [21] N. Magid, "Prevalence of Hydatid Cyst among Slaughter Animal in Duhok Province," M.Sc. Thesis, College of Veterinary Medicine, Duhok University, 2008.
- [22] R. A. S. Al-Nakeeb, "Seroepidemiological and Therapeutic Study on Hydatid Cyst Infection in Kirkuk and Tikrit Provinces," M.Sc. Thesis, College of Medicine, University of Tikrit, 2004.
- [23] A. M. Abdullah, "Epidemiological, Comparative Enzymatic and Total Protein content of Hydatid cyst of *Ecchinococcus granulosus* Isolated from Sheep and Goats in Duhok Province, Kurdistan Region of Iraq," M.Sc. Thesis, College of Education, University of Duhok, 2010.
- [24] K. S. A. Almufty, "Validity of Serological Tests in the Diagnosis of Hydatidosis," MSc. Thesis, College of Medicine, University of Duhok, 2012.
- [25] H. R. Rahimi, E. B. Kia, S. H. Mirhendi, A. Talebi, H. M. Fasihi, N. Jalali-zand and M. F. Rokni, "A new primer pair in ITS1 region for molecular studies on *Echinococcus granulosus*," *Journal of Public Health*, vol. 36, pp. 45- 49, 2007.
- [26] J. Bowles, D. Blair and D. P. McManus, "Genetic variants within the genus *Echinococcus* identified by mitochondrial DNA sequencing," *Molecular and Biochemical Parasitology*, vol. 54, pp. 165-173, 1992.
- [27] A. A. Hama, Z. I. Hassan, W. M. S. Mero, M. Interisano, B. Boufana and A. Casulli, "A morphologically unusual *Echinococcus granulosus* (G1 Genotype) cyst in a cow from Kurdistan - Iraq," *Epidemiology (Sunnyvale), Special Issue*, vol. 4, pp. 1, 2015.
- [28] M. R. Nejad, M. Roshani, F. Lahmi and N. E. Mojarad, "Evaluation of four DNA extraction methods for the detection of *Echinococcus granulosus* genotype," *Journal of Gastroenterology and Hepatology*, vol. 4, no. 2, pp. 91- 94, 2011.
- [29] M. Okamoto, M. Nakao, Y. Sako and A. Ito, "Molecular variation of *Taenia solium* in the world. Southeast Asian," *Journal of Tropical Medicine and Public Health*, vol. 32, pp. 90-93, 2001.
- [30] A. Dinkel, M. E. Njoroge, A. Zimmermann, M. Walz, E. Zeyhel, E. Elmahdi, I. U. Mackenstedt and T. Romig, "A PCR system for detection of species and genotypes of the *Echinococcus granulosus*-complex, with reference to the epidemiological situation in eastern Africa," *Journal of Parasitology*, vol. 34, pp. 645-653, 2004.
- [31] N. Ahmadi and A. Dalimi, "Molecular characterization of *Echinococcus granulosus* isolated from sheep and camel in Iran," *Archives of Razi Institute*, vol. 53, pp. 51-60, 2002.
- [32] D. P. McManus and R. C. A. Thompson, "Molecular epidemiology of cystic Echinococcosis," *Journal of Parasitology*, vol. 127, pp. 37-51, 2003.
- [33] A. A. Pour, S. H. Hosseini and P. Shayan, "Comparative genotyping of *Echinococcus granulosus* infecting buffalo in Iran using *co1* gene," *Journal of Parasitology Research*, vol. 108, pp. 1229-1234, 2001.
- [34] M. Nakao, D. P. McManus, P. M. Schantz, P. S. Craig and A. Ito. "A molecular phylogeny of the genus *Echinococcus* inferred from complete mitochondrial genomes," *Journal of Parasitology*, vol. 134, pp. 713- 722, 2007.
- [35] A. Obwaller, R. Schneider, J. Walochnik, B. Gollackner, A. Deutz, K. Janitschke, H. Aspöck and H. Auer, "*Echinococcus granulosus* strain differentiation based on sequence heterogeneity in mitochondrial genes of cytochrome oxidase-1 and NADH dehydrogenase-1," *Journal of Parasitology*, vol. 128, pp. 569-575, 2004.
- [36] Z. I. Hassan, A. A. Meerkhan, B. Boufana, A. A. Hama, B. D. Ahmed, W. M. S. Mero, S. Orsten, M. Interisano, E. Pozio and A. Casulli, "Two haplotype clusters of *Echinococcus granulosus sensu stricto* in northern Iraq (Kurdistan region) support the hypothesis of a parasite cradle in the Middle East," *Acta Tropica*, vol. 172, pp. 201-207, 2017